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Development of markers for detection of *Fusarium oxysporum* f. spp. *melonis* and *niveum*

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Introduction

- *Fusarium oxysporum* f.sp. *melonis* and f.sp. *niveum* are pathogens of melon and watermelon respectively.

- Seed-transmitted
Long term consequences of spread (wider rotation, soil disinfection, grafting)
- Moderate interest in the seed industry for seed testing (Nakt offers test)
- Proper control in seed production?
 - Dry heat treatment of the seeds?
 - Fungicide coating of seed? [benzimidazoles]

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Benefits of the PCR tool

- Replacing pathogenicity assay

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Ultimately for use in multiplex detection of pathogens

- Tool for a future all-inclusive PCR for cucurbits
 - *A. citrulli*
 - *P. s. lachrymans/peponis*
 - *Didymella bryoniae*
 - *Fusarium oxysporum*
 - Viruses (CGMMV, MNSV, SqMV)
 - ?
- *Fusarium solani* f.sp. *cucurbitae* out of scope; *F.oxysporum* considered to be of higher concern

Pathogenicity assays

- Lengthy (several weeks), labour intensive; hard to control / reproduce...esp. for watermelon
- But accurate strain characterization prerequisite for marker development



Catch 22:
no good pathogenicity assay without a good marker;
no good marker without a good pathogenicity assay

Published markers

Paper	Target	PCR ID	f.sp.	validation
Zhang et al. (2005) FEMS Microbiology Letters 249 39-47	rDNA, ITS	Fn-1/Fn-2 (SYBR Green)	niveum	7 Chinese strains, large outgroup
Lin et al. (2010) New Biotechnology 27, 4	Anon. (RAPD)	Fon-1/Fon-2 (gel-based)	niveum	Taiwanese isolates, 5 ATCC strains from outside of Taiwan
López-Mondéjar et al. (2012) Crop Protection 33 1-6	Anon. ("SCAR")	FOX S/FOX R (Taqman)	melonis	7 Spanish strains; cross-reaction with other f.spp.
Lievens et al. (2009) FEMS Microbiol. Lett. 300 (2), 201-215	SIX (effector gene)		lycopersici	to be done for melon! (niveum?)

SIX6 (continued)

- Lievens et al. 2009.
Of the 7 *Fol* genes homologues only found in several formae speciales for SIX6

SIX6 Pos	Six6 Neg
<i>lycopersici</i> (many)	<i>radicis lycopersici</i> (many)
<i>melonis</i> (1)	<i>niveum</i> (3) (2/4) <small>ENR CSIRO Plant Pathol. 69 (2): 232</small>
<i>radicis cucumerinum</i> (23/28)	<i>radicis cucumerinum</i> (5/28)
<i>niveum</i> (2/4) <small>ENR CSIRO Plant Pathol. 69 (2): 232-243 (2011)</small>	<i>cucumerinum</i> (many)
<i>vasinfectum</i> (Australian) (19/2 VCGs) <small>ENR CSIRO Plant Pathol. 69 (2): 232-243 (2011)</small>	<i>conglutinens</i> (1) <i>cubense</i> (3) <i>asparegi</i> (3)
<i>passiflorae</i> (3) <small>ENR CSIRO Plant Pathol. 69 (2): 232-243 (2011)</small>	<i>dianthi</i> (2), <i>gladioli</i> (3), <i>lili</i> (2) <i>luffae</i> (2), <i>opuntiarum</i> (3), <i>spinaciae</i> (3), <i>tulipae</i> (3)

SIX6

- 568 bp fragment, primers designed by Harrie Koenraad
- Lievens et al. (2009) different primers, 973 bp amplicon
- Lievens, B., Houterman, P. and Rep, M. Effector gene screening allows unambiguous identification of *Fusarium oxysporum* f. sp. *lycopersici* races and discrimination from other formae speciales FEMS Microbiol. Lett. 300 (2), 201-215 (2009)
- **SIX6 diagnostic test for Austr. isolates of *F. o. f. sp. vasinfectum***
Chakrabarti, A., Rep, M., Wang, B., Ashton, A., Dodds, P. and Ellis, J. Variation in potential effector genes distinguishing Australian and non-Australian isolates of the cotton wilt pathogen *Fusarium oxysporum* f. sp. *vasinfectum* Plant Pathol. 60 (2), 232-243 (2011)

Strains for (further) validation of candidate markers

- "AFLP collection"
 - f. sp. *melonis*: 4 AFLP groups: C (12), D (1), F (6), H (1) $\Sigma=20$
 - f. sp. *niveum*: 5 AFLP groups: B (4), C (1), E (14), F (2), G (1) $\Sigma=21$
- Isolates from commercial melons in Southern Europe
 - 53 f. sp. *melonis*
 - 14 non-pathogenic *F. oxysporum*
- Non-pathogenic *Fusarium oxysporum* from melon seeds
- *Fusarium oxysporum* f. sp. *niveum* being characterized
- Collection strains

Pathogenicity determination

- Pathogenicity testing on fully susceptible assay plant
- Strains of all known races of f.sp. melon included

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Results Fn1-Fn2 primers

- Many non-pathogenic *F. oxysporum* strains react with Fn-1/Fn-2 PCR
- This was observed by Lin et al. in their study as well
- Validation of Fn-1/Fn2 PCR will no longer be pursued
- Multicopy target attractive



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Results AFLP collection with 3 PCRs

AFLP group	# total	# FOX S / FOX R	# SIX6	# Fon-1 / Fon-2
<i>F. o. melonis</i>				
C	12	12	11	0
D	1	1	1	0
F	6	0	6	0
H	1	0	1	0
<i>F. o. niveum</i>				
B	4	0	0	0
C	1	1*	1**	0
E	14	0	12	14
F	2**	0	1**	1**
G	1	0	0	0

* Original strain from CBS negative for both
 ** Same strain is negative for both SIX6 and Fon-1/Fon-2

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Result "melon collection" plants Southern Europe and seed isolates

Source and Identity	# total	# FOX S / FOX R	# SIX6	# Fon-1 / Fon-2
Plants				
<i>F. o. melonis</i>	53	51	53	ND?
<i>F. oxysporum</i>	14	1	0	ND?
Seed				
<i>F. oxysporum</i>	112	1	0	ND?

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SIX6 (+ FOX S/FOX R) promising for melonis, but all diversity covered?

- Kerry O'Donnell et al. (2009)
A two-locus DNA sequence database for typing plant and human pathogens within the *Fusarium oxysporum* species complex
Fungal Genetics and Biology 46, 936–948
- 850 isolates typed, 101 EF-1a, 203 IGS rDNA, and 256 two-locus sequence types (STs)
- *melonis* 18 isolates, 5 haplotypes (STs)
- *niveum* 5 isolates, 4 haplotypes (STs)
- 3 *melonis*, 4 *niveum* DNA preps received
 - one *melonis* DNA prep negative with SIX6; corresponds to CBS strain

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F. oxysporum f.sp. *melonis* CBS strains

- All SIX6 negative (and FOX S FOX R negative)
- All non-pathogenic....
- Four SIX1 positive

ST ^a	NRRL				
165	38472 CBS 420 80	<i>F. oxysporum</i> f. sp. <i>melonis</i> race 0		Cucumis melo	Israel
167	38473 CBS 421 90	<i>F. oxysporum</i> f. sp. <i>melonis</i> race 0		Cucumis melo	Israel
165	38474 CBS 422 80	<i>F. oxysporum</i> f. sp. <i>melonis</i> race 0		Cucumis melo	Israel
167	38475 CBS 423 90	<i>F. oxysporum</i> f. sp. <i>melonis</i> race 2	ICMPP 10416	Cucumis melo	Israel
167	38478 CBS 424 80	<i>F. oxysporum</i> f. sp. <i>melonis</i> race 1,2		Cucumis melo	Israel

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Pathogenicity assay CBS strains (De Ruiter Seeds)



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Primers for SIX1 targeting conserved region

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2891 ATGTAACCTTGGGGCTTC      2894 GCGTATGTCAGAGCTGC
Query 1 ATGGGCTATAGCAGGACCTTGGGGCTTCATAGTCTGGTTTGGCTTACTTAAAGGCTAGG
AABR1000429 2132 .....
AABR1001117 111 .....
AABR1001528 1785 .....
AABR1001571 630 .....
AABR1001175 2891 .....

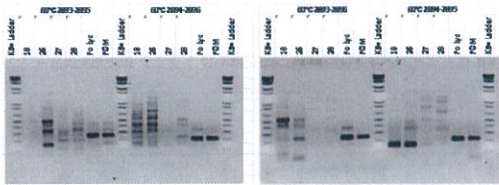
2894 GAGCTATGAGGGCTGGTC      2895 GAGGGGCTGTGAGATTGTCA
Query 234 GAGTG-GCAGGAGCTATGAGGGCTGGTCACATTGCAANTTAGAAAGGTAGCTTTGGGAG 300
AABR1000429 1899 .....
AABR1001117 111 .....
AABR1001528 1552 .....
AABR1001571 866 .....
AABR1001175 2625 .....
    
```

Query: F. o. f.sp. lycopersici SIX1
AGHE01001175: F. o. f.sp. melonis

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SIX1 primer screening of F. o. f.sp. niveum (SIX6 neg.)



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Conclusions and discussion

- SIX6 is a very promising candidate marker for *F. oxysporum* f.sp. *melonis* (confirmed by M. Rep, unpublished)
- FOX S / FOX R PCR does not pick up *F. oxysporum* f.sp. *melonis* belonging to two AFLP groups, but may have additional value for SIX6 negatives in other groups (one strain)
- Is SIX6 unstable if *F. oxysporum* f.sp. *melonis* (and *niveum*) is propagated outside the host? Or is additional marker (SIX1) needed?
- Fon-1 / Fon-2 and SIX6 reactions are correlated in *F. oxysporum* f.sp. *niveum* AFLP groups, but *F. oxysporum* f.sp. *niveum* group(s) appear to exist that lack both markers.

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Conclusions and discussion

- Are SIX genes present in non-pathogenic *F. oxysporum*? (False-positives.)
- A conclusion on acceptability of marker will have to account for some risk: pathogen/non-pathogen distinction (assay plant treatment, assay plant type, incubation conditions/humidity, effect of strain culturing on media); collection always limited.
- But balanced by benefits of screening larger numbers of seed/suspects.

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Schmidt et al. *BMC Genomics* 2013, **14**:119

- While some *ff. spp.* are monophyletic, others are composed of several clonal lineages that appear to have independently acquired the ability to infect the same host plant [4-6]. This polyphyletic origin was likely caused by horizontal transfer of chromosomes encoding host specific virulence genes between *Fo* lineages, thereby allowing the distinction of members of a *f. sp.*, not by overall genetic relatedness, but by the presence or absence of certain LS chromosomes [1].

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